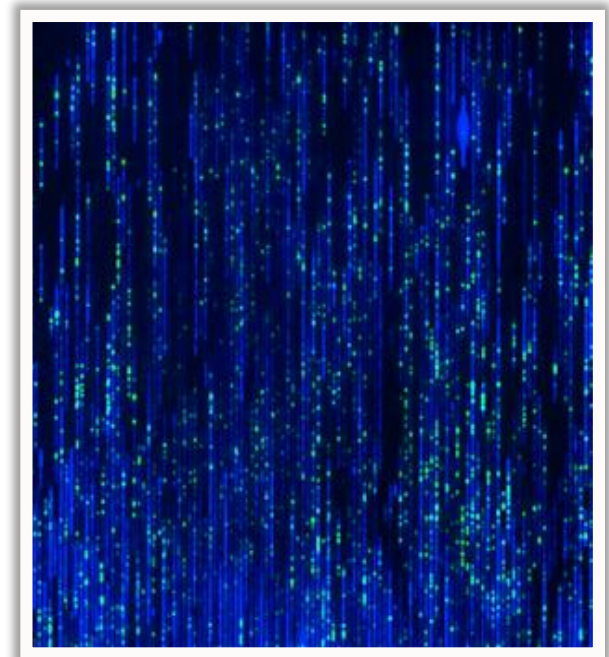


# Optical mapping at the French Plant Genomic Center with the BioNano Irys system

Céline Jeziorski  
HeliOr meeting  
November 23th, Cordoba



# The French Plant Genomic Center

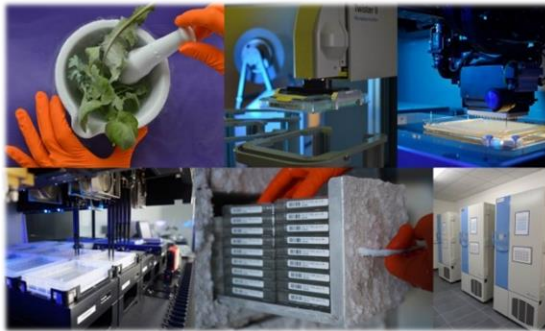


ISO 9001:2008  
Octobre 2005

Created in 2004 by INRA (French National Institute for Agricultural Research)

- **Depository of genomic libraries for the scientific community**

⇒ BAC libraries



# The French Plant Genomic Center

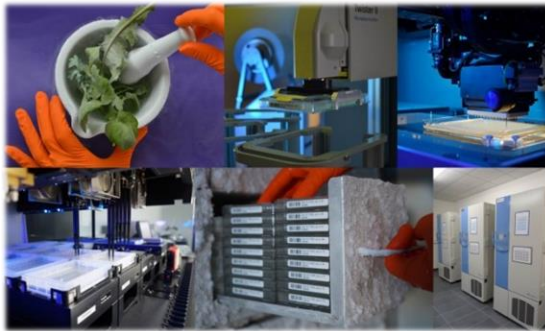


ISO 9001:2008  
Octobre 2005

Created in 2004 by INRA (French National Institute for Agricultural Research)

• **Depository of genomic libraries for the scientific community**

⇒ BAC libraries



• **A dedicated structure to assist plant genomic programs**

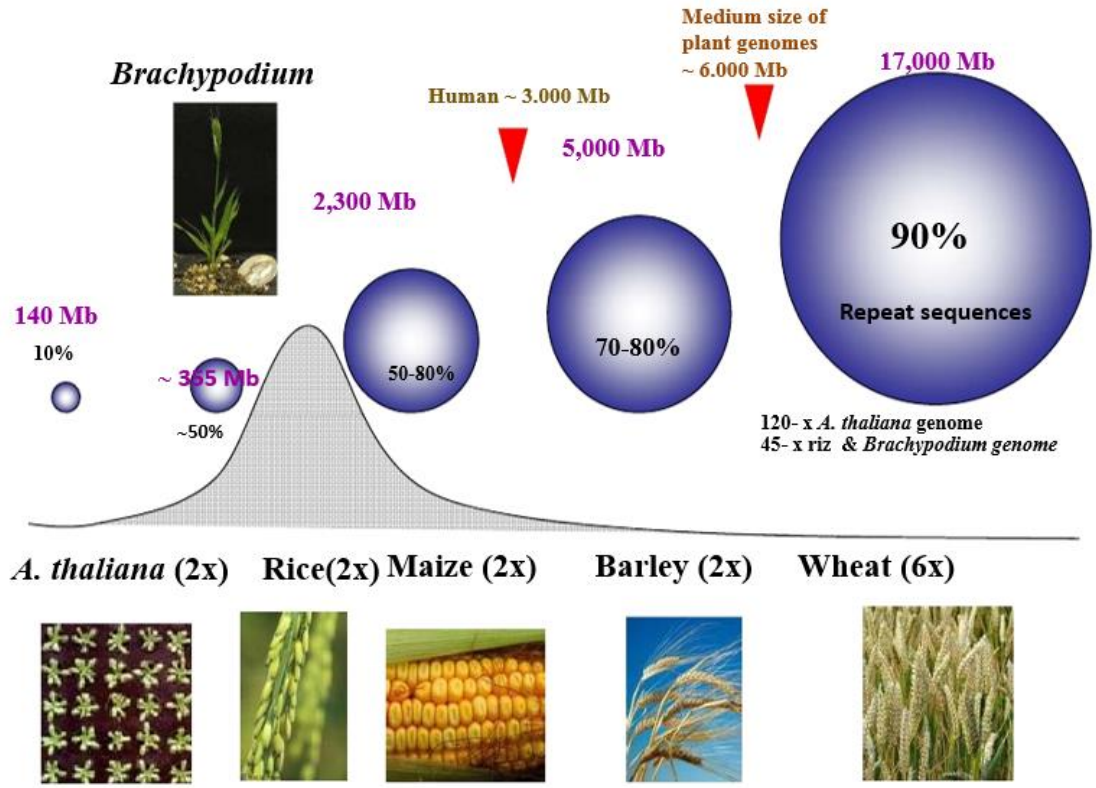
- ⇒ Distribute the genomic resources at the international level
- ⇒ Provide high quality research material and efficient tools to study plant genomes
- ⇒ Develop innovative solutions
- ⇒ Develop genomic projects in collaboration
- ⇒ Host scientists in the frame of collaborations

## Interactions with laboratories around the world



- **3 233 242 BAC clones** distributed during the last 5 years (2011-2015)

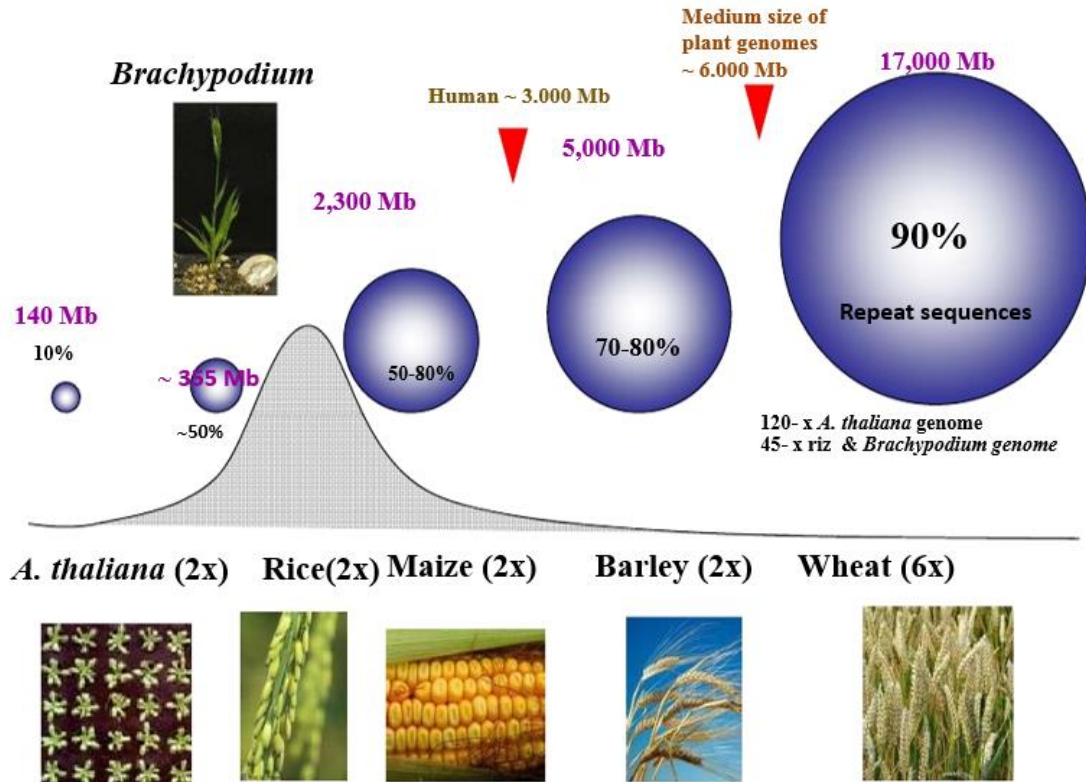
# The complexity of plant genomes



Plant genomes are challenging:

- Genome size
- Repeat elements (>80% in maize, barley, sunflower...)
- Polyploidy

# The complexity of plant genomes



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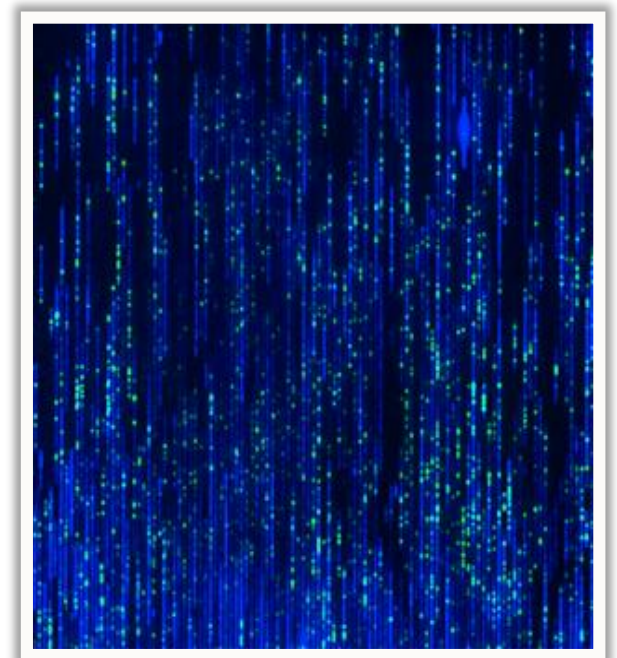
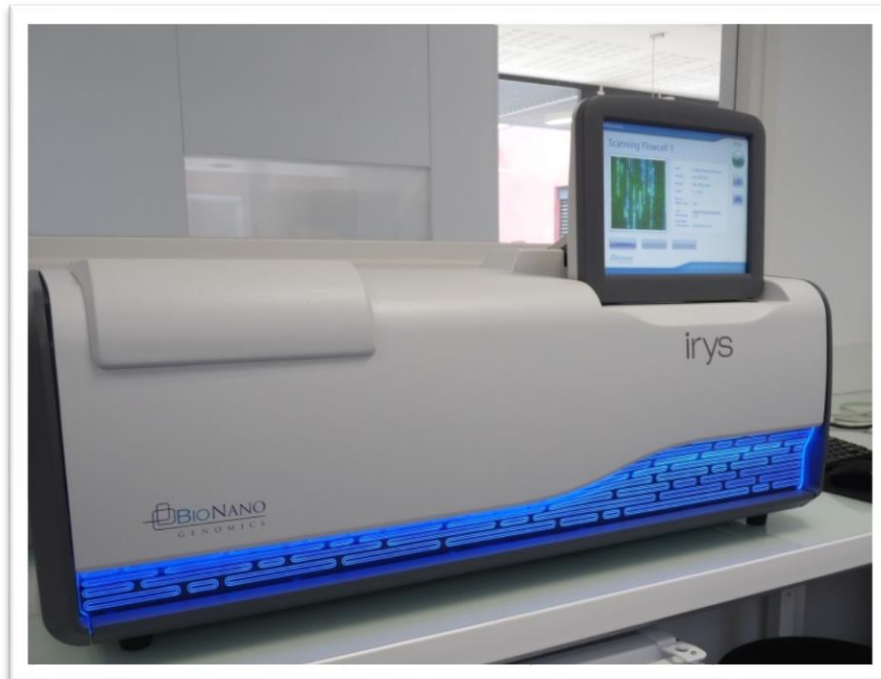
The use of BAC libraries is important

- Reduce the complexity
- Focus on specific target/markers
- Manage the diversity

The Bionano Irys system: another tool to better study the complex genomes

Advantages of BioNano optical mapping:

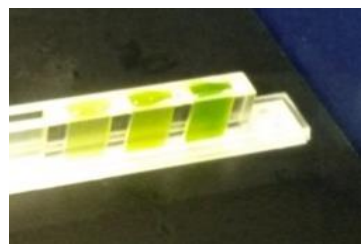
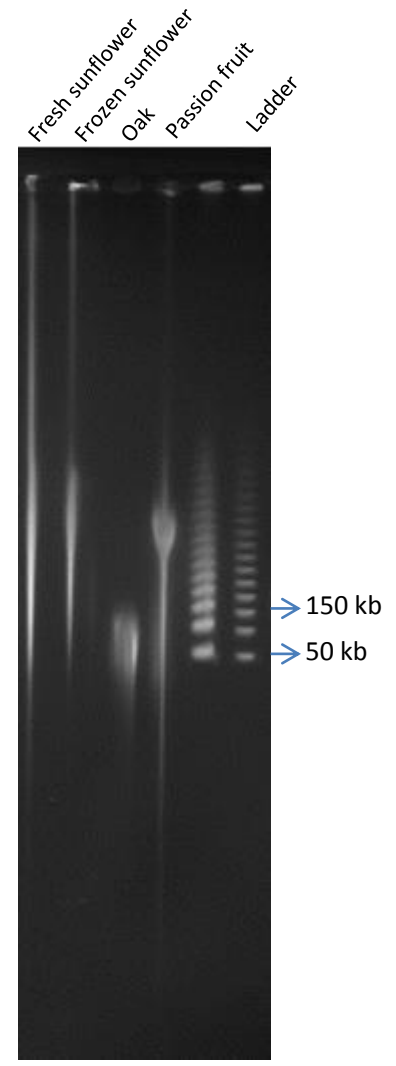
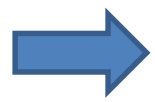
- Direct visualization of long DNA molecules (>100 kb)
- Physical structure and map
- Provides real physical distance information



# Isolation of High Molecular Weight DNA



Using Bionano Plant DNA Isolation Kit

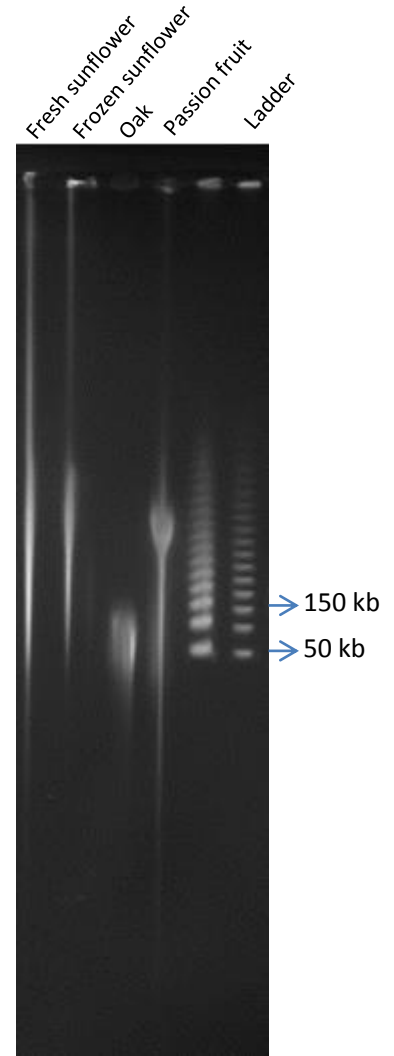




# Isolation of High Molecular Weight DNA

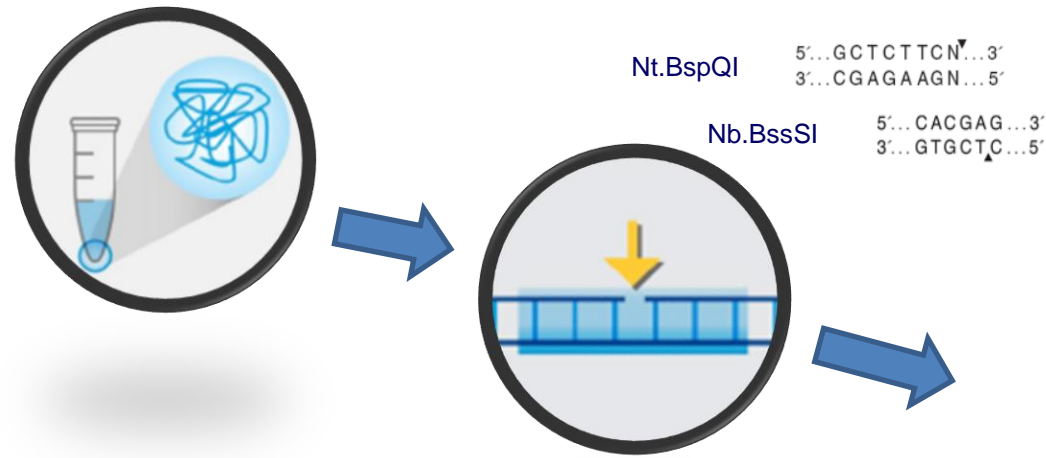


Using Bionano Plant DNA Isolation Kit



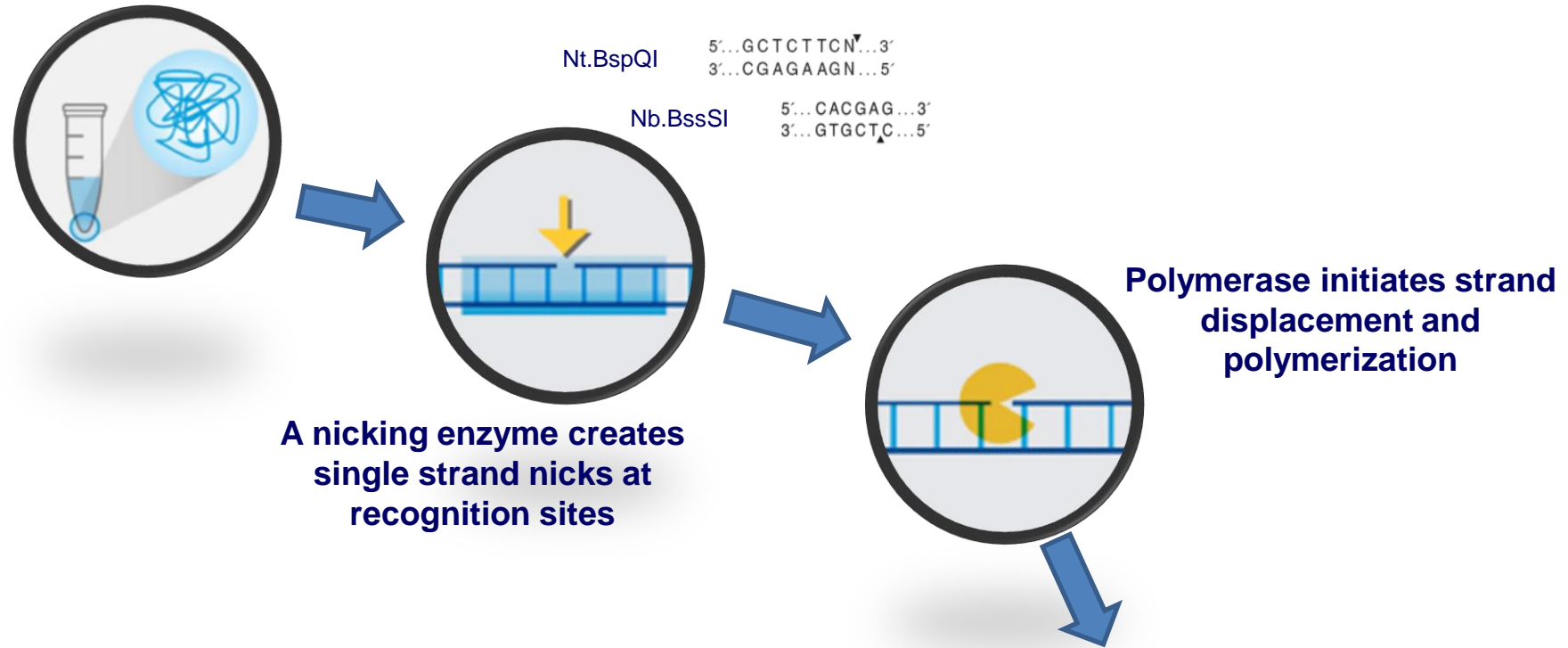
Important to have a high quality of HMW DNA

# Nick Label Repair Stain (NLRs) workflow

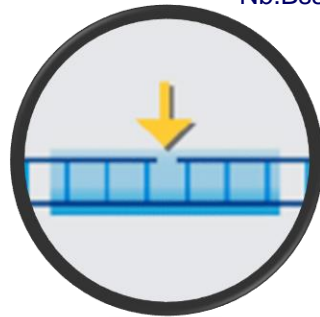


**A nicking enzyme creates  
single strand nicks at  
recognition sites**

# Nick Label Repair Stain (NLRs) workflow



# Nick Label Repair Stain (NLRs) workflow



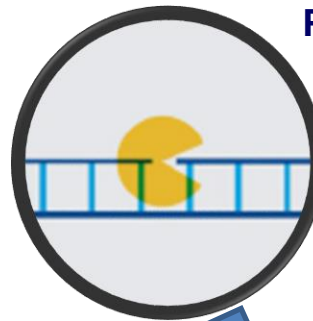
**A nicking enzyme creates single strand nicks at recognition sites**

Nt.BspQI

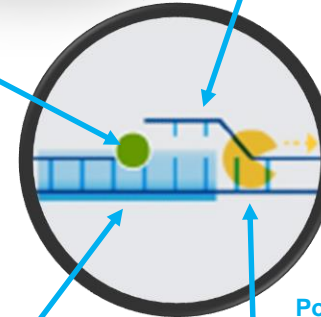
5'...GCTCTTCN<sup>v</sup>...3'  
3'...CGAGAAGN...5'

Nb.BssSI

5'...CACGAG...3'  
3'...GTGCTC...5'



**Polymerase initiates strand displacement and polymerization**



**Fluorescent nucleotides are incorporated to label enzyme recognition sites**

Fluorescent nucleotides

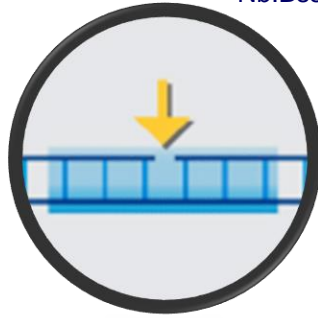
DNA displaced Strand

Polymerase

Nick Site



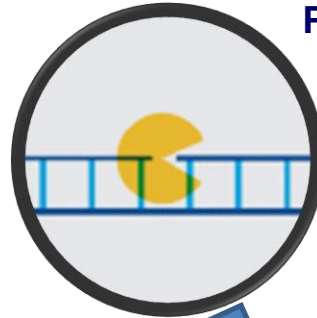
# Nick Label Repair Stain (NLRS) workflow



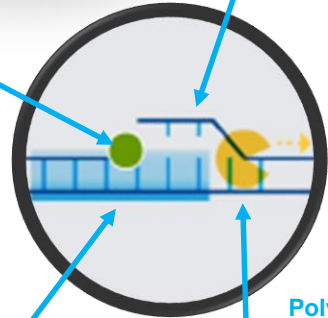
**A nicking enzyme creates single strand nicks at recognition sites**

Nt.BspQI 5'...GCTCTTCN<sup>v</sup>...3'  
3'...CGAGAAGN...5'

Nb.BssSI 5'...CACGAG...3'  
3'...GTGCTC...5'



**Polymerase initiates strand displacement and polymerization**



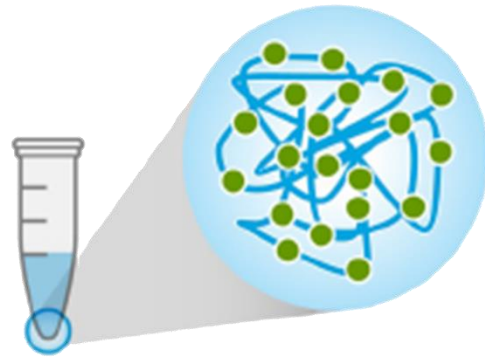
**Fluorescent nucleotides are incorporated to label enzyme recognition sites**

Fluorescent nucleotides

DNA displaced Strand

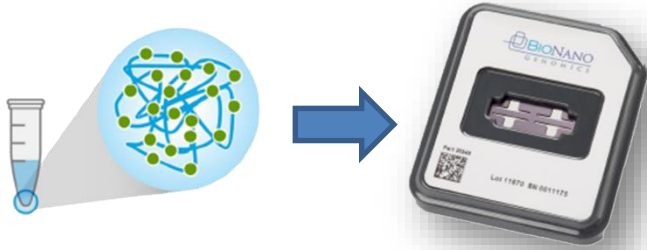
Nick Site

Polymerase



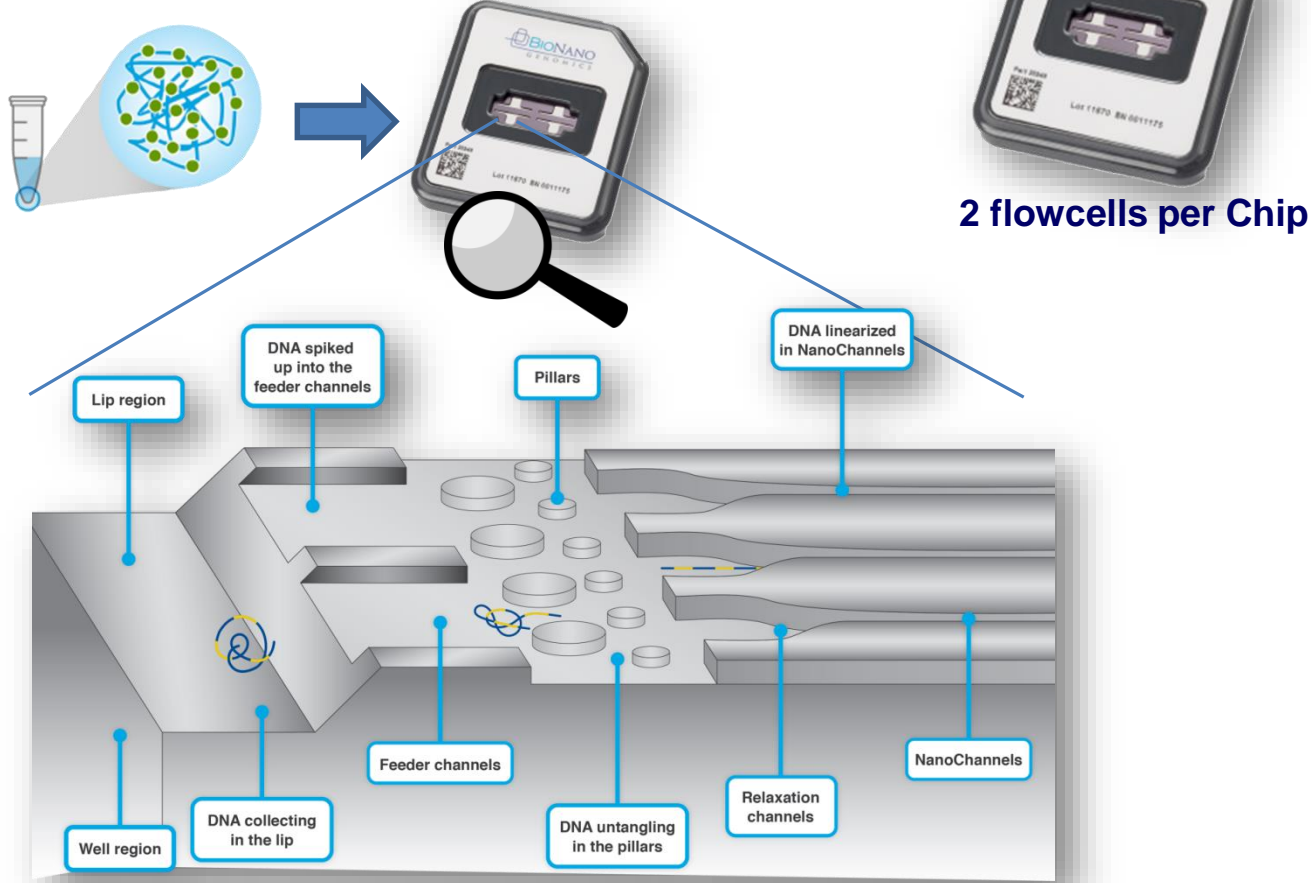
**The nicked-labeled DNA is stained with YOYO-1**

The nicked-labeled-stained  
DNA is loaded on a chip



# Irys System technology

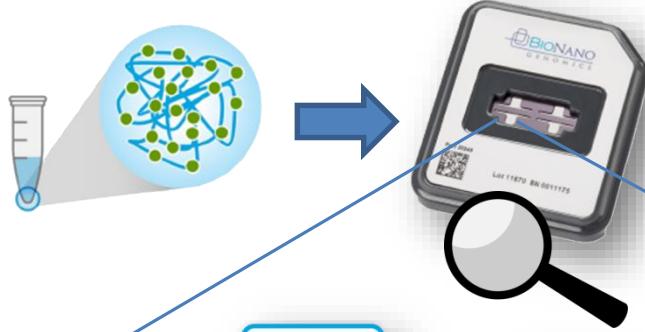
The nicked-labeled-stained DNA is loaded on a chip



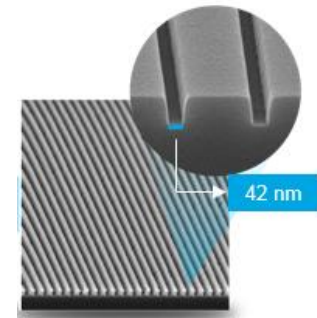
DNA molecules are linearized in the NanoChannels

# Irys System technology

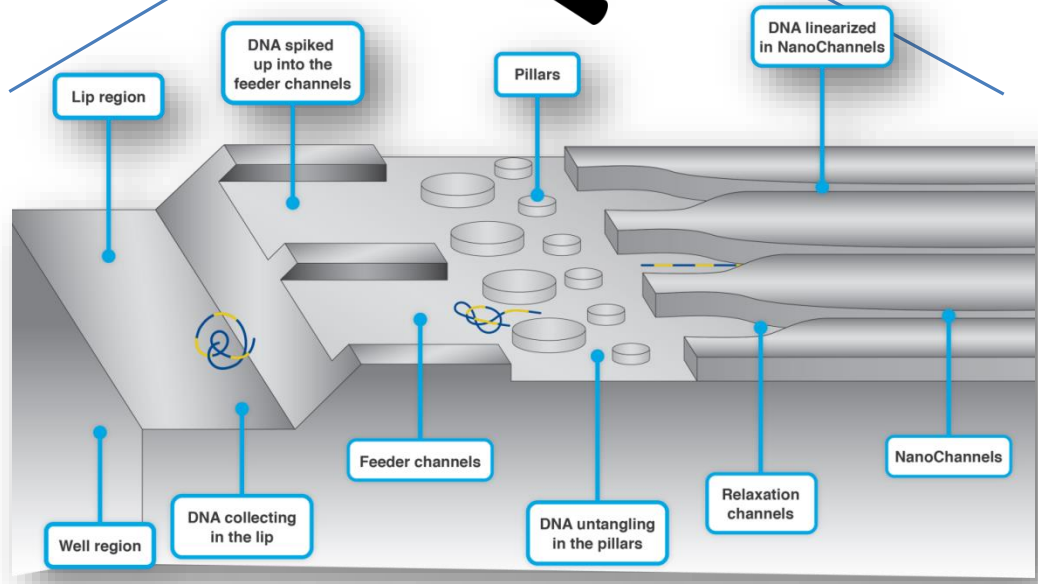
The nicked-labeled-stained DNA is loaded on a chip



2 flowcells per Chip



13 000 channels per flowcell

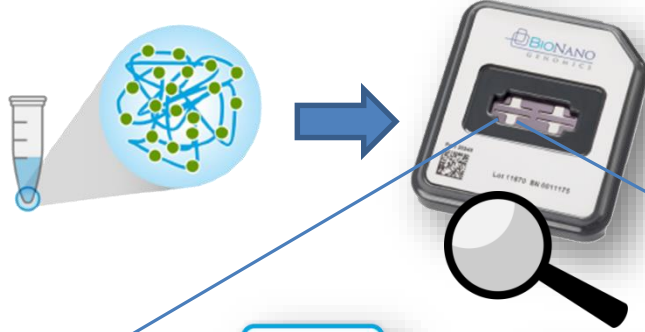


DNA molecules are linearized in the NanoChannels

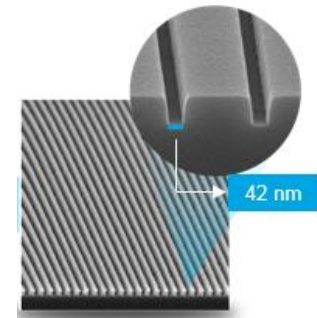


# Irys System technology

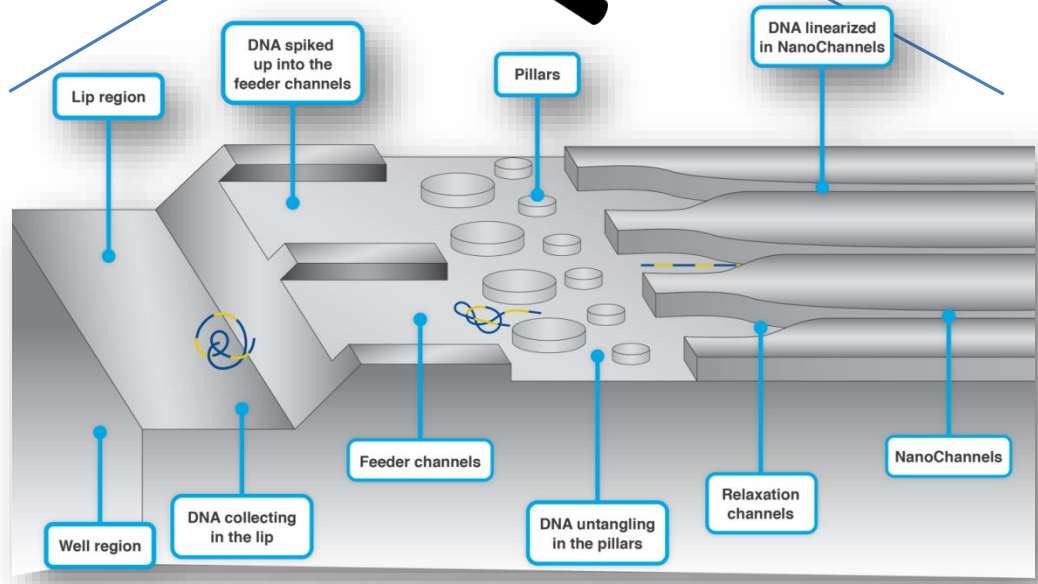
The nicked-labeled-stained DNA is loaded on a chip



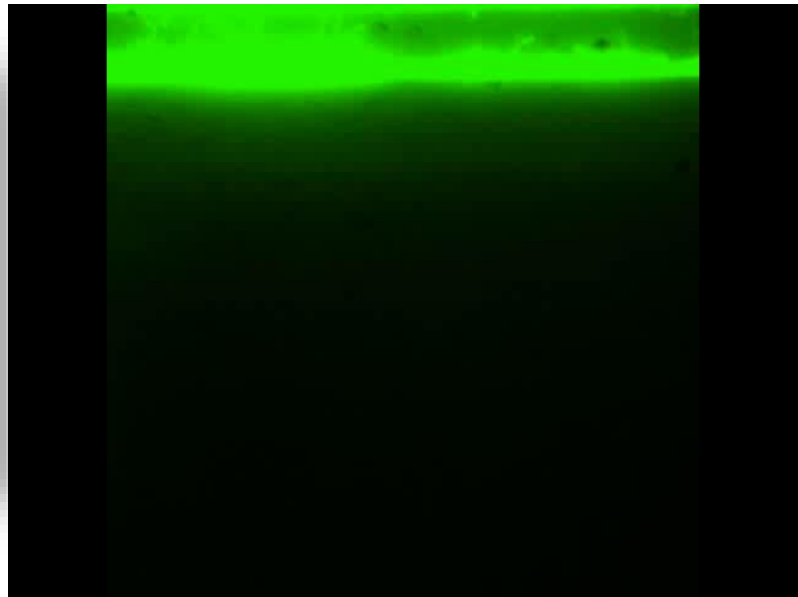
2 flowcells per Chip



13 000 channels per flowcell

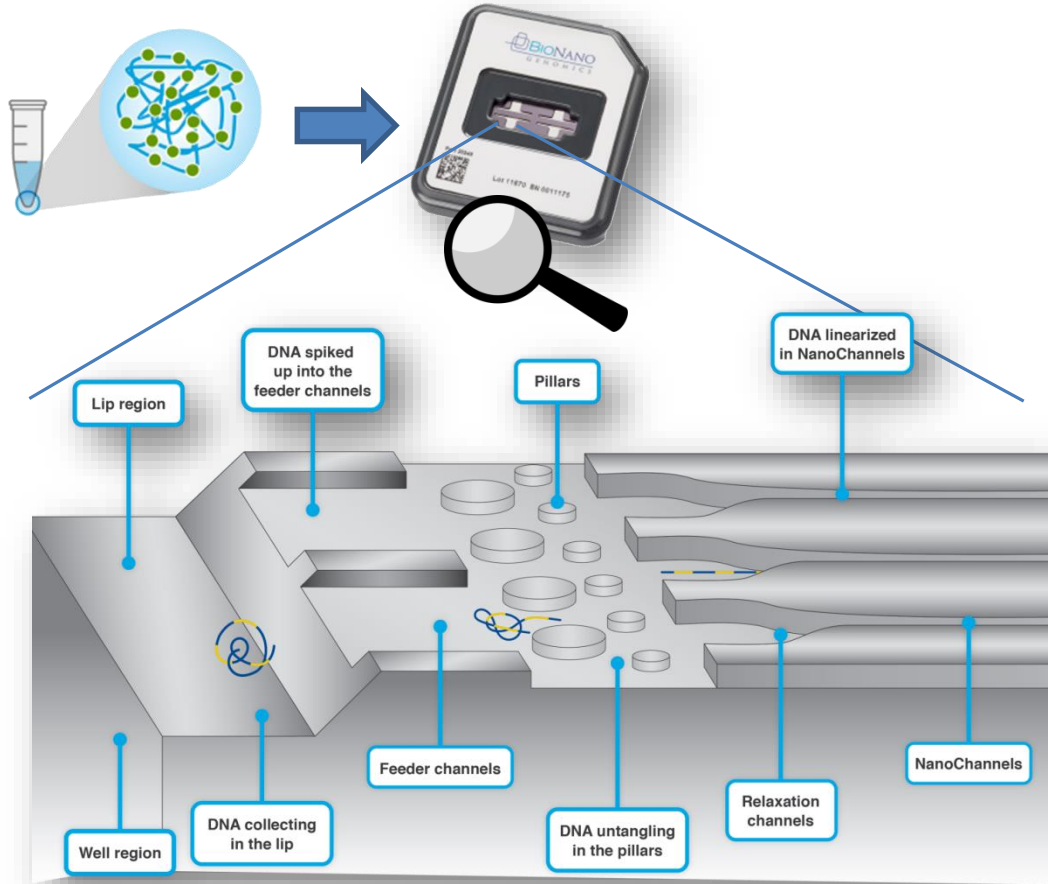


DNA molecules are linearized in the NanoChannels

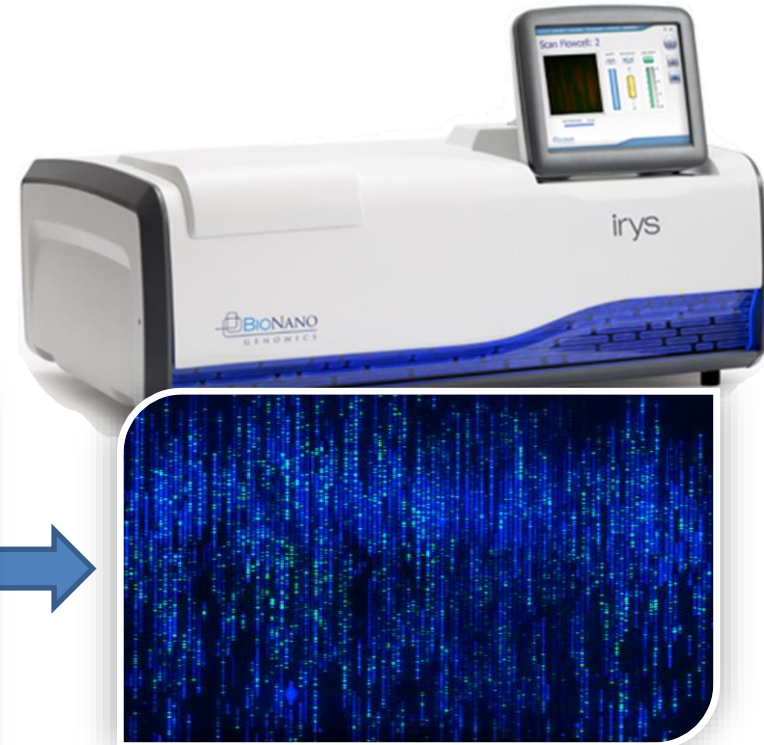


# Irys System technology

The nicked-labeled-stained DNA is loaded on a chip



DNA molecules are linearized in the NanoChannels



The Irys System collects images of linearized DNA molecules

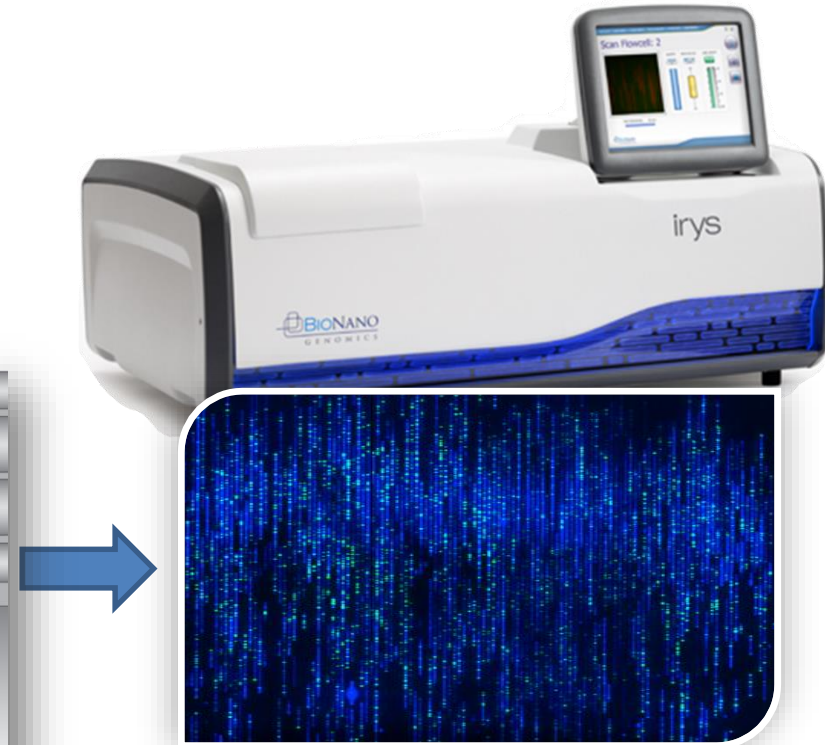
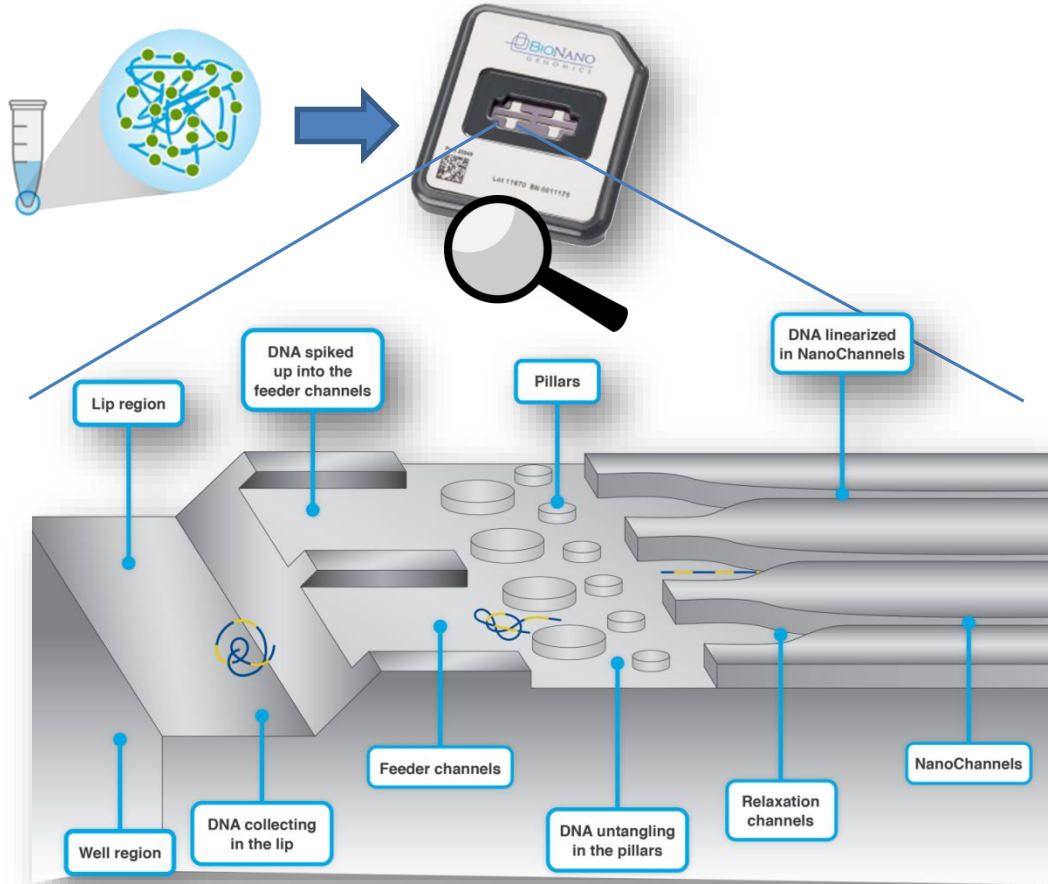
# Irys System technology

The nicked-labeled-stained DNA is loaded on a chip

30 scans/run

24h-30h/run

50-100 Gb raw data / run/ flowcell

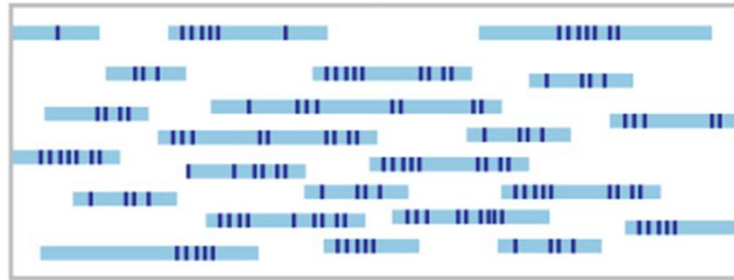
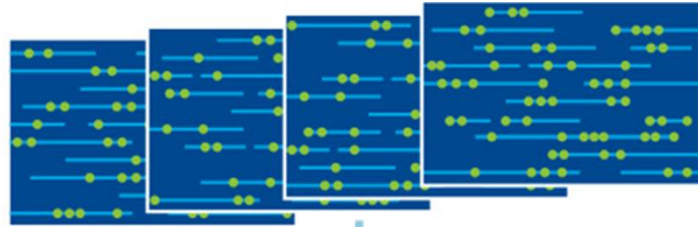
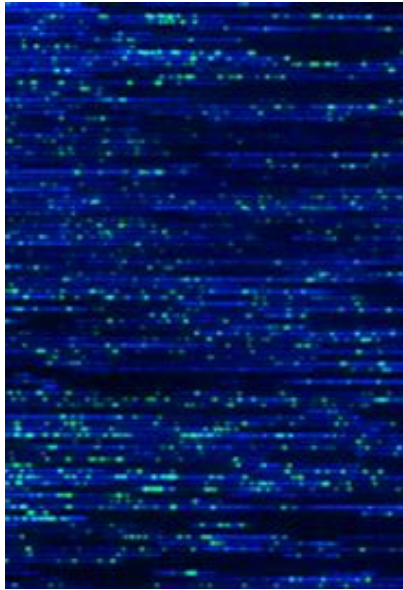


DNA molecules are linearized in the NanoChannels

The Irys System collects images of linearized DNA molecules

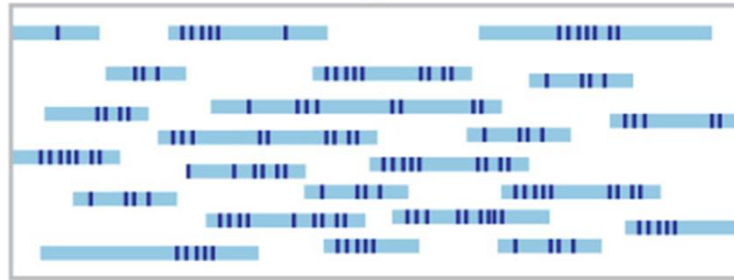
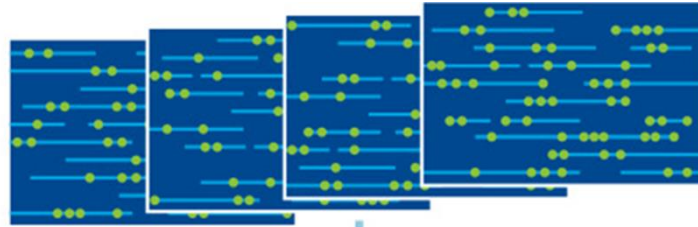
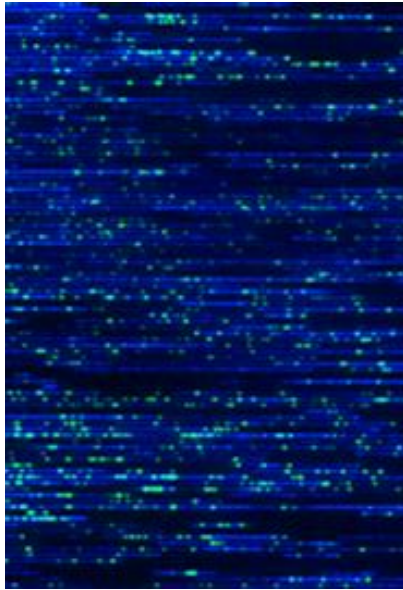
# Data Analysis

Algorithms convert images  
into digital molecules



# Data Analysis

Algorithms convert images  
into digital molecules



*De Novo* assembly is made to  
construct a consensus genome map

## Genome Assembly and Hybrid Scaffolding



Ex: Xiao et al. 2015

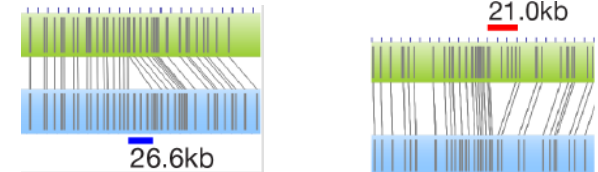
- Enhance & refine genome (references and draft genomes)
- Orient and align sequencing reads using a hybrid scaffold
- Close the contig gaps
- Combined with sequencing reads for *de novo* assembly in species without a reference genome

## Long range SV detection and discovery (2Kb to >1Mb)



Ex: Cao et al. 2014

- In/Dels –Inversions
- Translocations
- CNVs

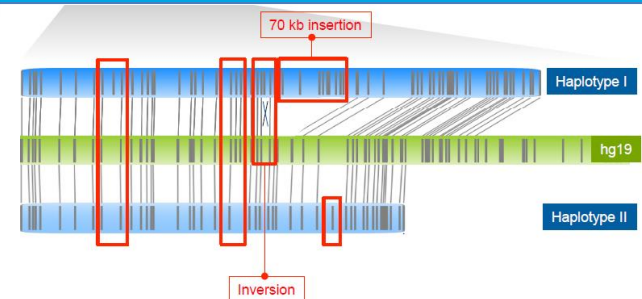


## Haplotyping



Ex: Dong et al. 2016

- Comparison of 2 haplotypes with a reference



## Examples of optical maps:

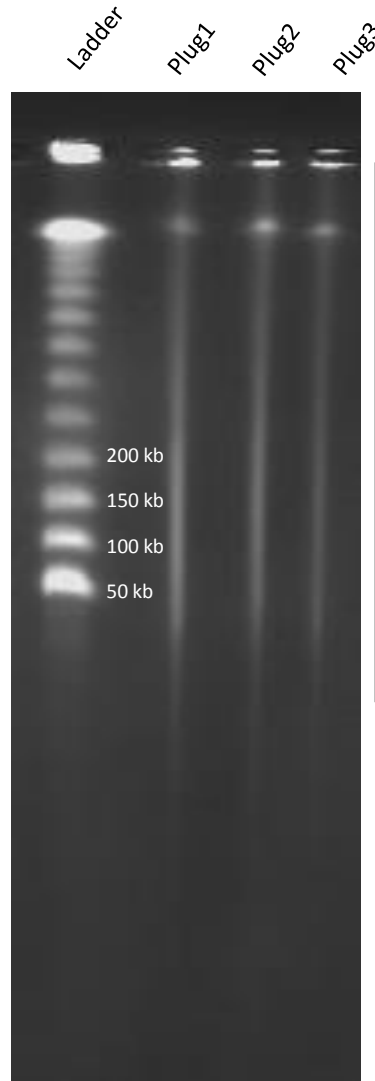
**Sunflower (*Helianthus annuus*) XRQ  
and**

**Sunflower broomrape (*Orobanche cumana*) IN-23**

# Example of *H. annuus* XRQ



**4 g of fresh young leaves**  
**3 days of dark treatment**  
**2 nicking enzymes (BspQ1 & BssS1)**



### Scanning Flowcell 2

User: Nathalie Rodde

Sample: 2016\_07\_11\_Han\_XRQ\_BssS1...

Recipe: 2016\_07\_11\_han\_xrq\_bsss1\_fc2

Cycle: 26 of 30

Pause After Cycle: N/A

Time Left (hr): 05:35

Estimated Completion: 12/07/2016 17:30

**BspQ1**

7,2 labels/100kb

**1100 Gb raw data**

**300X**

**N50 168 kb**

**BssS1**

17,2 labels/100kb

**983 Gb raw data**

**270X**

**N50 148 kb**



# Example of *H. annuus* XRQ

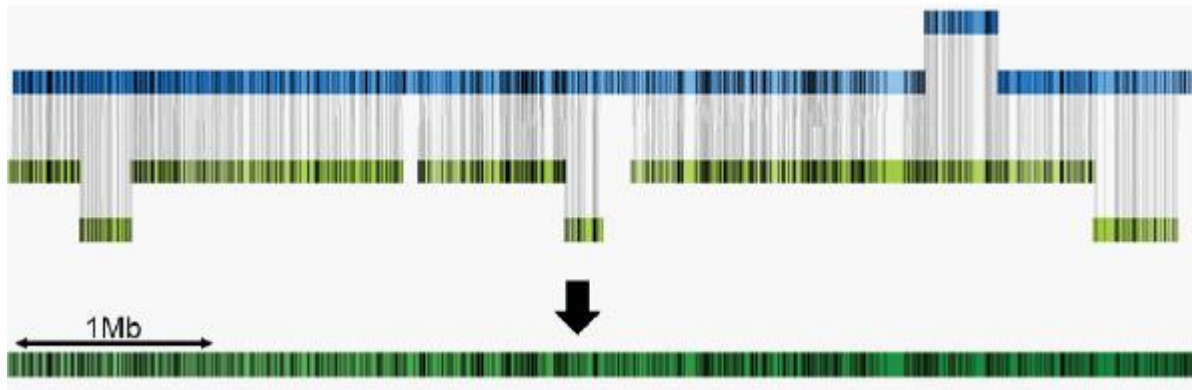
<b>BspQ1 XRQ</b>	<b>PacBio assembly</b>	<b>BioNano <i>de Novo</i> assembly</b>	<b>Hybrid scaffolding (+ not scaffolded PacBio contigs)</b>
Number of contigs	11676	2959	9433
N50 length (Mb)	0.521	1.444	2.195
Genome map length (Gb)	2.9	3.2	3.6

**Estimated genome size: 3.6 Gb**

**BioNano Map**

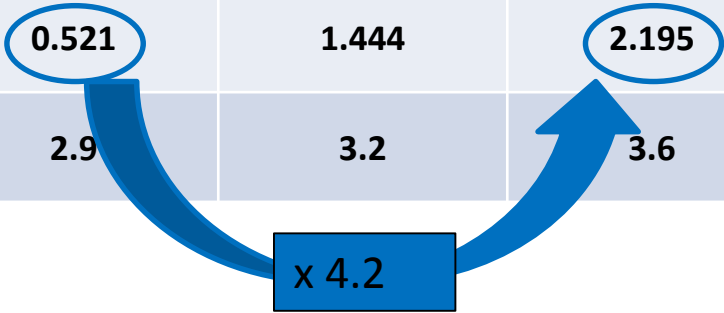
**PacBio assembly**

**Hybrid scaffolding**



# Example of *H. annuus* XRQ

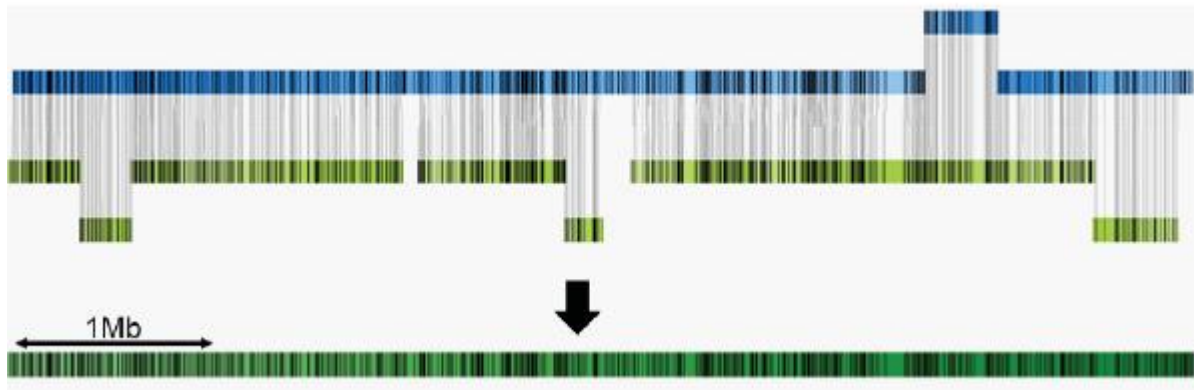
BspQ1 XRQ	PacBio assembly	BioNano <i>de Novo</i> assembly	Hybrid scaffolding (+ not scaffolded PacBio contigs)
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**BioNano Map**

**PacBio assembly**

**Hybrid scaffolding**

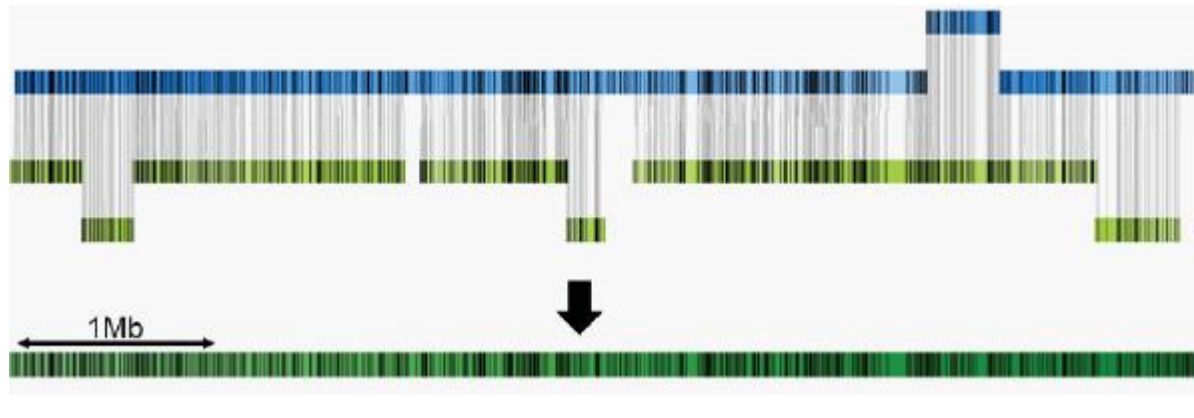


Work on BssS1 data still in progress

BioNano Map

PacBio  
assembly

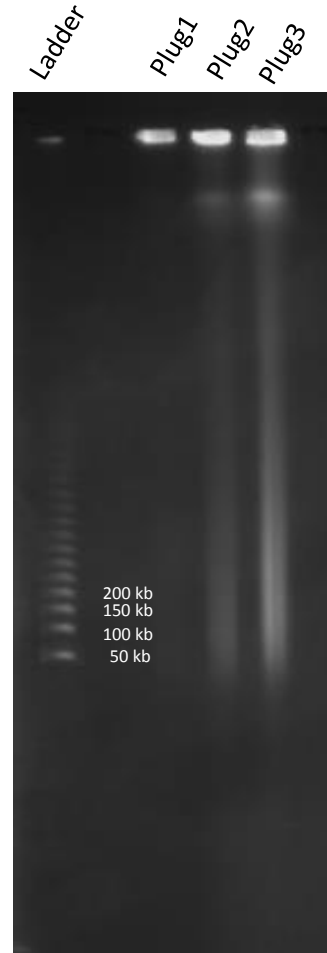
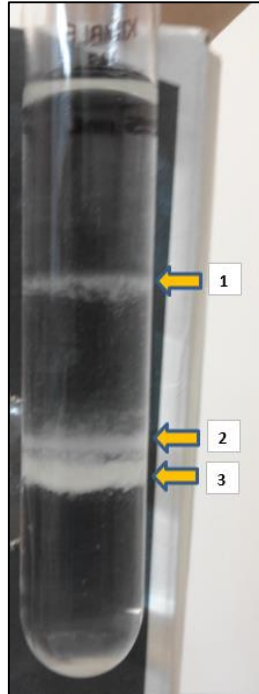
Hybrid  
scaffolding



# Example of *O. cumana* IN-23

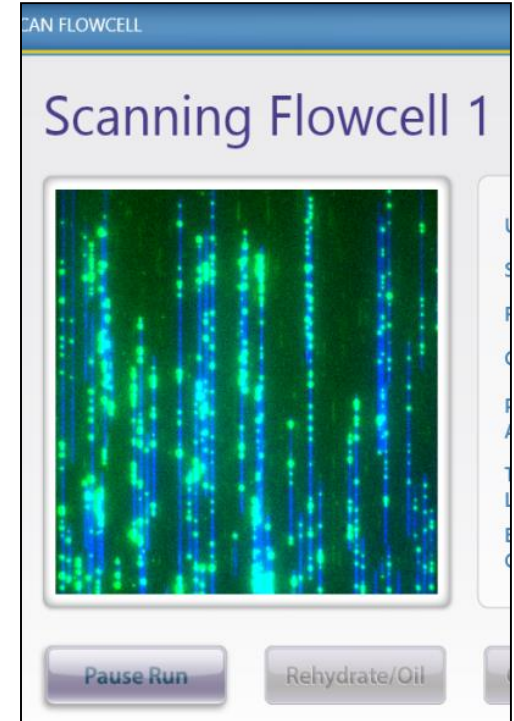


2 g of frozen plant  
 Extraction optimization (sugar)  
 1 nicking enzyme (BspQ1)



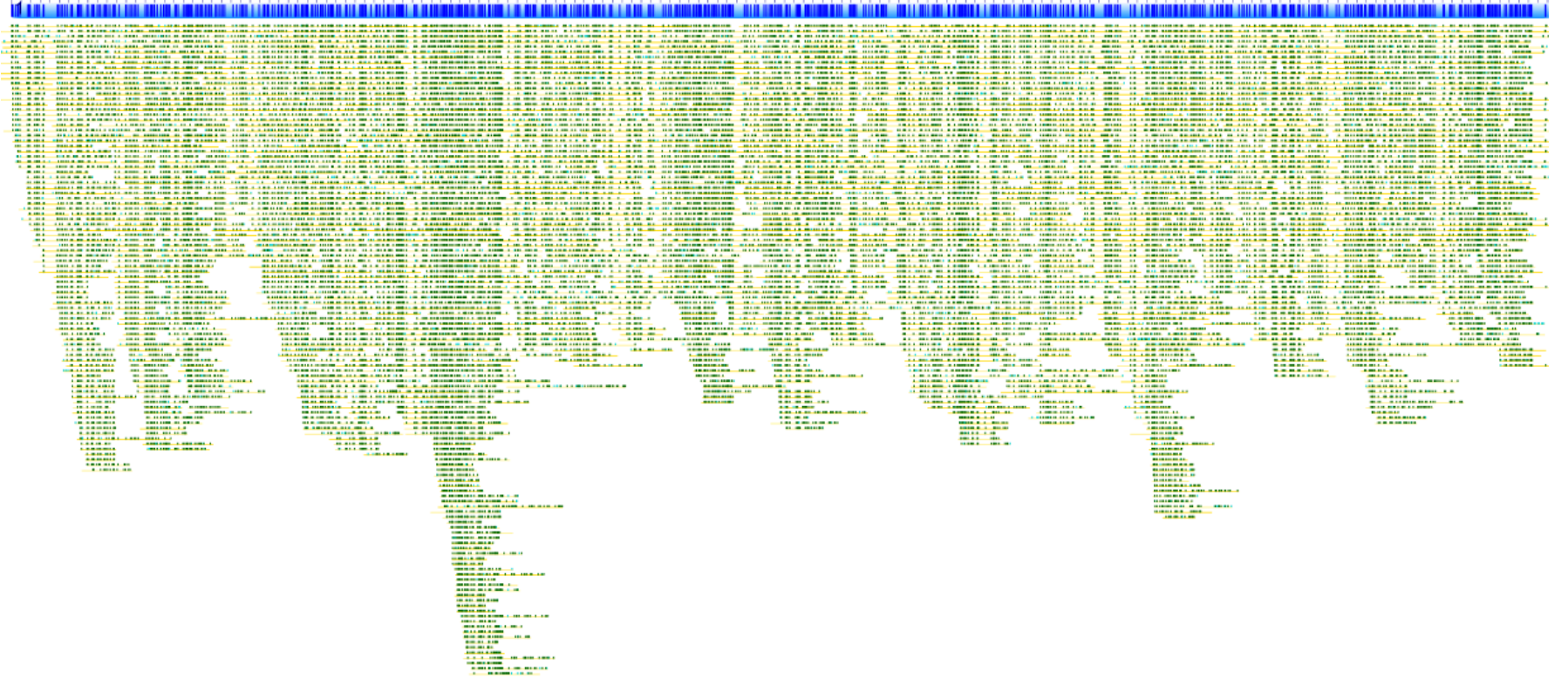
**BspQ1**  
 11,5 labels/100kb

**680 Gb raw data**  
**370X**  
**N50 143 kb**



# Example of a contig assembly of *O.cumana* IN-23

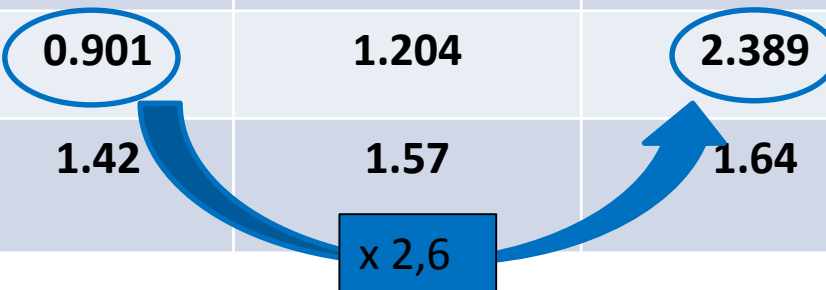
0 0.2M 0.4M 0.6M 0.8M 1M 1.2M 1.4M 1.6M 1.8M 2M 2.2M 2.4M 2.6M 2.8M 3M 3.2M 3.4M 3.6M 3.8M 4M 4.2M 4.4M 4.6M 4.8M 5M 5.2M 5.4M 5.6M 5.8M 6M 6.2M 6.4M



<b>BspQ1 IN-23</b>	<b>Pacbio assembly</b>	<b>Bionano <i>de novo</i> assembly</b>	<b>Hybrid scaffolding (+ not scaffolded Pacbio contigs)</b>
<b>Number of Contigs</b>	<b>7199</b>	<b>1901</b>	<b>6405</b>
<b>N50 lenght (Mb)</b>	<b>0.901</b>	<b>1.204</b>	<b>2.389</b>
<b>Genome Maps Size (Gb)</b>	<b>1.42</b>	<b>1.57</b>	<b>1.64</b>

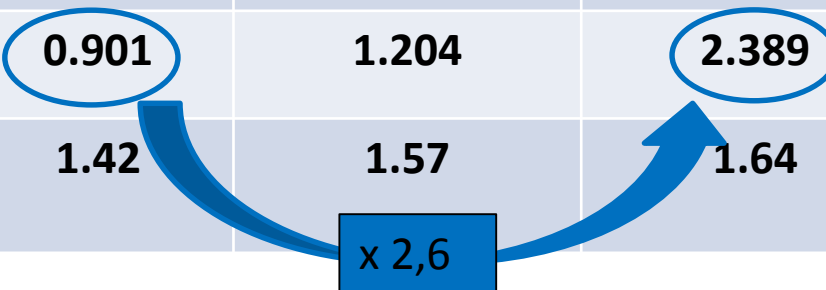
**Estimated genome size: 1.4 Gb – 2 Gb**

<b>BspQ1 IN-23</b>	<b>Pacbio assembly</b>	<b>Bionano <i>de novo</i> assembly</b>	<b>Hybrid scaffolding (+ not scaffolded Pacbio contigs)</b>
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## Next steps:

- More data to improve the quality of assembly (selection of higher molecular weight DNA molecules)
- Optical map with other enzyme BssS1 (19,4 labels/100kb)
- 2 steps hybrid scaffolding



# Conclusion

- **CNRGV offers genomic tools to better understand the complex genomes**
  - **Construction of BAC libraries**
  - **Construction of optical maps**
- **The optical map approach can be useful for genome assembly/finishing (hybrid scaffolding and 2 steps hybrid scaffolding), and also allows to detect structural variations**
- **Other applications can be developed on the machine (dual labeling)**

# Thank you for your attention



Céline JEZIORSKI  
 Arnaud BELLEC  
 Sonia VAUTRIN  
 Genséric BEYDON  
 Nathalie RODDE  
 William MARANDE  
 Joëlle FOURMENT  
 Elisa PRAT  
 Nadine GAUTIER  
 Nadège ARNAL  
 Céline CHANTRY-DARMON  
 Stéphane CAUET  
 David PUJOL  
 Laetitia HOARAU  
 Hélène BERGES



Jérôme GOUZY  
 Marie-Claude BONIFACE  
 Johann LOUARN  
 Nicolas POUILLY  
 Nicolas LANGLADE  
 Stéphane MUNOS



<http://cnrgv.toulouse.inra.fr/>

